



Tehran University of Medical
Sciences Publication
<http://tums.ac.ir>

Iran J Parasitol

Open access Journal at
<http://ijpa.tums.ac.ir>



Iranian Society of Parasitology
<http://isp.tums.ac.ir>

Original Article

Evaluation of Immunogenicity of Cocktail DNA Vaccine Containing Plasmids Encoding Complete GRA5, SAG1, and ROP2 Antigens of *Toxoplasma gondii* in BALB/C Mice

Razi NASERIFAR¹, *Fatemeh GHAFARIFAR², Abdolhosein DALIMI², Zohreh SHARIFI³, Kavous SOLHJOO⁴, Kami HOSSEINIAN KHOSROSHAHI²

1. Department of Medical Parasitology, Faculty of Medicine, Ilam University of Medical Sciences, Ilam, Iran
2. Department of Parasitology and Entomology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran
3. Blood Transfusion Research Center, High Institute for Research and Education in Transfusion Medicine, Tehran, Iran
4. Department of Medical Microbiology, Faculty of Medicine, Jahrom University of Medical Sciences, Jahrom, Iran

Received 10 Jan 2015

Accepted 19 Aug 2015

Keywords:

Toxoplasma gondii,
Cocktail DNA Vaccine,
pcGRA5,
pcSAG1,
pcROP2

***Correspondence**

Email:
ghafarif@modares.ac.ir

Abstract

Background: Severe and fatal complications of toxoplasmosis urge development of effective vaccines against the disease. The current study was performed to evaluate cocktail DNA vaccine containing plasmids encoding GRA5, SAG1, and ROP2 genes of *Toxoplasma gondii* in BALB/c mice in Tarbiat Modares University in 2012.

Methods: The plasmids containing complete GRA5, SAG1, and ROP2 genes were mass extracted and then the recombinant plasmids were administered via intramuscular injections according to immunized mice three times with three-week intervals. Then splenocytes were cultured, and proliferation as well as cytokine assays were carried out. The other mice in each group were inoculated by the parasite and mortality of the mice was evaluated on a daily basis.

Results: The results of cytokine assay for INF- γ were higher in the mice that received the cocktail DNA containing recombinant plasmids. Evaluation of proliferation of splenocytes using the MTT (3-[4,5-dimethylthiazol-2-yl]-2,5 diphenyl tetrazolium bromide) assay indicated induction of cellular response. Measurement of total IgG and the isotypes of IgG1 and IgG2a showed that the cocktail DNA stimulated IgG and IgG2a production in comparison with the control groups ($P<0.05$). Furthermore, the survival rate of mice in the groups that received the cocktail DNA was significantly higher than that in the control groups ($P<0.05$).

Conclusion: Administration of the cocktail DNA vaccine led to production of higher levels of IFN- γ , confirmed by secretion of IgG2a, and the immune response was shifted toward Th1. Thus, the cocktail DNA containing the recombinant plasmids can be an appropriate candidate for immunization against toxoplasmosis.

Introduction

Contamination with *Toxoplasma gondii* is one of the commonest parasitic infections in human and other vertebrates (1). There is an increasing tendency towards not using antiparasitic drugs. The high-tech DNA vaccines have proposed a good perspective for development of cocktail vaccines (2-3).

In recent years, considerable achievements have been made on identification of an appropriate vaccine. Most of these attempts have focused on surface antigens. Among the antigens, SAG1 is one of the major candidates for vaccine development (2). Secretory antigens of *Toxoplasma* have an important role in stimulation of the protective immune system. These antigens are expressed by cystic bradyzoites as well as tachyzoites (4). The major components of secretory antigens are GRA molecules known as vaccine candidates (5). ROP2 antigen, expressed by tachyzoites, bradyzoites, and sporozoites, are considered as candidates for development of vaccines against *T. gondii* (6). Rhoptry proteins are associated with various cellular components and have different functions. ROP2 (54 kDa) is one of the main rhoptry proteins in the *T. gondii* (7).

SAG1 is the main surface protein of the tachyzoites and comprises 3-5% of the total protein volume of the tachyzoites. SAG1 is a tachyzoite stage-specific protein, which is present only in tachyzoites and cannot be found in sporozoites and bradyzoites. SAG1 shows the highest immunogenicity in *T. gondii* tachyzoites thus, has been employed in development of cocktail DNA vaccines against toxoplasmosis (8-12).

Another group of secretory antigens is GRA, among which GRA5 is one of the main antigens of *Toxoplasma* secreted in different life stages of the parasite. If the whole antigen is used, it can be employed in development of effective test for detection of antibodies against toxoplasmosis (13-14).

The aim of this study was to evaluate the expression of plasmids encoding complete GRA5, SAG1, and ROP2 gens of *T. gondii* in eukaryotic CHO cells and immunogenicity of cocktail DNA vaccine containing these plasmids in BALB/c mice.

Materials and Methods

This research was carried out in Department of Parasitology, Faculty of Medicine, Tarbiat Modares University, Tehran, Iran in 2012 for evaluating the Cocktail DNA vaccine containing plasmids encoding complete GRA5, SAG1, and ROP2 antigens of *T. gondii* in BALB/C mice.

Preparation of recombinant plasmid

The construction and expression of pcDNA3 plasmids (Invitrogen, USA) contain complete genes of ROP2 gene (1,686 bp), SAG1 (960 bp) and GRA5 (363 bp) were formerly prepared and expressed in CHO cells (15-17).

The primers used for each gene were as follows:

ROP2: The primers of *T. gondii* ROP2 were designed according to the gene sequence obtained from GenBank, with the accession No. Z36906. The PCR product size was 1686 bp. Forward primer, 32nt 5' ATT AAG CTT ATG GAA AAC TGT GCG TCG GTC AG 3' Hind III Reverse primer 29 nt; 5' ATT GAA TTC TCA TGC CGG TTG TCC ATC AG 3' EcoRI

SAG1: The sequence of SAG1 gene of *T. gondii* RH strain (complete code: 960 bp) was obtained from GenBank, with the accession No. AY217784, and the following primers were designed.

Forward primer, 27nt 5'-ATT AAG CTT ATG TTT CCG AAG GCA GTG-3' Hind III Reverse primer, 26nt 5'-ATT GAA TTC TCA CGC GAC ACA AGC TG-3' EcoRI

GRA5: The sequence of GRA5 gene of *T. gondii* RH strain (complete code: 363 bp) was obtained from Gen Bank, with the accession No. EU918733. The size of PCR product was 363 bp.

Forward primer 25nt, 5'- AAG CTT ATG GCG TCT GTA AAA CGC G - 3Hind III Reverse primer 27nt, 5'- GAA TTC TTA CTC TTC CTC GGC AAC TTC - 3'EcoR I. The recombinant plasmids were transformed into *E. coli*, strain TG1. Following mass replication of the bacterium were extracted from the bacteria using a plasmid extraction kit (Qiagen, Germany).

Animals

Female BALB/c mice at the age of 6-8 wk and bodyweight of 20 ± 1 g were purchased from the Razi Vaccine and Serum Research Institute. The animals were kept under standard conditions of light, food, and water. The study was approved by the Ethics Committee of Tarbiat Modares University, with number 52/75793 in 23-December-2010, Tehran, Iran.

Parasite

Tachyzoites of *T. gondii*, strain RH, obtained from the Department of Parasitology, Tarbiat Modares University. The antigen required for the ELISA was prepared from the parasite. Moreover, the parasites were used for stimulation of peripheral blood mononuclear cells to be used in MTT and cytokine assays. The antigens obtained were measured using a $0.2\ \mu\text{m}$ sterile filter (Nagle Company, USA), and the protein concentration was determined according to the Bradford method.

Immunization

Sixty female BALB/c mice at the age of 6-8 wk were divided into six groups, each including ten mice. Two groups were considered as control; one control group received $100\ \mu\text{l}$ BPS and the other control group received $75\ \mu\text{g}$ pcDNA3 at the volume of $100\ \mu\text{l}$. Of the remaining groups, groups 3-5 respectively received injections of $50\ \mu\text{g}$ of plasmids ($25\ \mu\text{g}$ pcSAG1 + $25\ \mu\text{g}$ pcROP2, $25\ \mu\text{g}$ pcSAG1 + $25\ \mu\text{g}$ pcGRA5 and $25\ \mu\text{g}$ pcROP2 + $25\ \mu\text{g}$ pcGRA5), accompanied by $25\ \mu\text{g}$ pcDNA3 at the volume of $100\ \mu\text{l}$. The last cocktail group received $75\ \mu\text{g}$ of a combination of the three plasmids ($25\ \mu\text{g}$ pcSAG1 + $25\ \mu\text{g}$ pcGRA5 + $25\ \mu\text{g}$ pcROP2) at equal ratio at the volume of

$100\ \mu\text{l}$. The injections were performed in bilateral biceps, and in each side, the maximum injection volume was $50\ \mu\text{l}$. The immunization was performed in three steps with the interval of three weeks.

Serological tests

To perform the serological tests, serum samples were obtained from the mice at days 42 and 63 post-immunization and were kept at $-20\ ^\circ\text{C}$ until measurements. Then, IgG, IgG1, and IgG2a were measured. The IgG subtypes were evaluated using the mouse monoclonal antibody isotyping reagents (Sigma, USA). To determine the total IgG level using ELISA, $20\ \mu\text{g}/\text{ml}$ *Toxoplasma* lysate antigens (TLA) was coated on the ELISA plate overnight. Then, after blocking and washing, the mouse sera was added to the wells at 1:10 concentration, and after washing again, anti-mouse antibody conjugated with HRP was added to the wells. As the incubation period was over, the plates were washed and TMB substrate was added. Twenty min later, reaction stopper solution, sulfuric acid 2M, was added, and the absorbance OD was read at the wavelength of 450 nm using an ELISA reader (Bio-Rad, USA). Determination of IgG1 and IgG2a were performed according to the manual of the kit (Mouse Monoclonal Antibody Isotyping Reagents, Sigma) using the ELISA method.

Evaluation of cellular immunity

Cellular immunity was evaluated by determination of lymphocyte proliferation and differentiation using the MTT and cytokine assay.

The MTT was used for evaluation of lymphocyte proliferation and differentiation. After performing the culture of peripheral blood mononuclear cells, cytokine assay was carried out for determination of IFN- γ and IL-4 levels. To this end, seven weeks after the last immunization, five mice from each group were killed and the lymphocytes were extracted from the spleen. The lymphocytes were cultured in the medium containing 20% FCS (Gibco) and RPMI 1640 (Gibco). In each

well, 3×10^5 cells were incubated for 72 h at 37 °C in the presence of 40 µg/ml of TLA and 5% CO₂ in triplicate. Then, the contents of the wells were centrifuged and until cytokine assay, the supernatant was frozen at -80 °C. Cytokine assay was performed using a kit (R&D Systems, Minneapolis, MN, USA).

To carry out the MIT assay, 20 µl of tetrazolium (Roche, Germany) at the concentration of 5 mg/ml was added to each well, and the samples were incubated at 37 °C for 4 h. Then, the plates were centrifuged at 1000 g for 10 min, the supernatant was removed, and 100 µl DMSO was added to each well. The OD values of the plates were determined at the wavelength of 540 nm.

Survival assay

To evaluate the survival rate of the mice, seven weeks after the last immunization, five mice in each group received an injection of 10^4 live tachyzoites of *T. gondii*, strain RH, and the mortality of the mice were evaluated on a daily basis.

Statistical analysis

For statistical analyses between different groups, at first the data were checked for normality distribution by Colmogorov-Smirnov

test. The analyses were performed by one way ANOVA and Mann-Whitney tests. The statistically significant results are the *P*-value less than 0.05.

Results

The recombinant plasmid extracted from bacteria and confirmed with PCR (Fig. 1).

Expression of pc-ROP2, pc-SAG1 and pc-GRA5 recombinant plasmids in CHO cells after 48 h of transfection in comparison with empty pc-DNA were confirmed by SDS-PAGE and Western blot (Fig. 2).

The lowest mean OD value for total IgG in the first and second blood samplings was related to the PBS group, while the highest values were obtained for the pcGRA5+ pcROP2 group (Table 1).

Furthermore, the cut-off value determined according to measurement of total IgG in serum samples of seronegative mice was (Mean + 3 × SD) 0.199 (Table 1).

In the first and second blood samplings, the lowest mean OD values related to pcGRA5+pcSAG1 and pcGRA5+pcROP2 groups, respectively (Table 2).

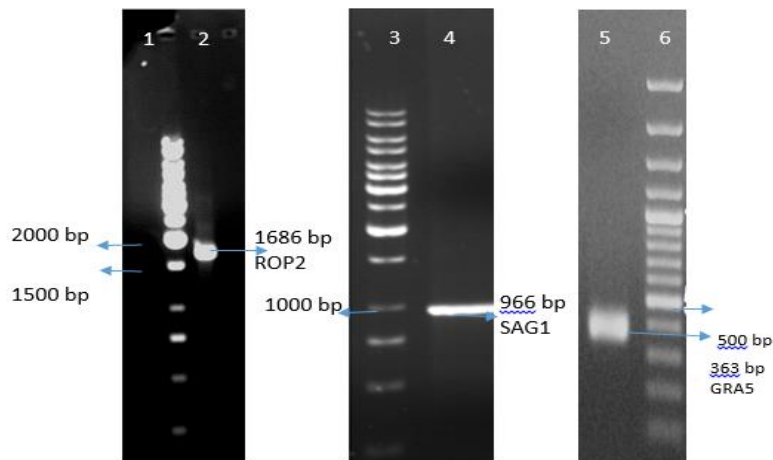


Fig. 1: ROP2, SAG1 and GRA5 PCR amplification and gel electrophoresis.

Lanes 1, 3 and 6 1 Kbp DNA ladder. Lane 2, PCR product of ROP2 (1686 bp); Lane 4, PCR product of SAG1 (966 bp); Lane 6, PCR product of GRA5 (363 bp)

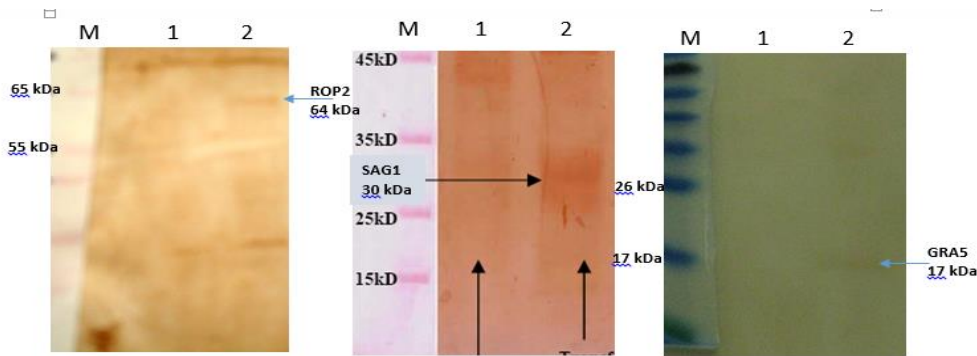


Fig. 2: Western blot of expressed genes in CHO cells. M, protein marker; Lanes 1, transfected cells containing pc-DNA 3 (negative control); Lines 2, Lane 6, transfected cells containing pc-ROP2, pc-SAG1 and pc-gra5

Table 1: Comparison of mean OD values for total IgG determined using ELISA test in serum samples of mice for two blood samplings

Number of group	Immunization regimen	Total IgG on day 42		Total IgG on day 63	
		Mean±SD	Sig ($P<0.05$)	Mean±SD	Sig ($P<0.05$)
1	pcSAG1+ pcROP2	0.263±0.02	(5, 6) *	0.317±0.02	(5, 6) *
2	pcGRA5+ pcSAG1	0.276±0.01	(5,6) *	0.310±0.01	(5, 6) *
3	pcGRA5+ pcROP2	0.269±0.01	(5,6) *	0.320±0.02	(5,6) *
4	pcGRA5+pcSAG1+ pcROP2	0.261±0.01	(5, 6) *	0.275±0.02	(5, 6) *
5	PBS	0.193±0.02	(1, 2, 3,4)*	0.171±0.01	(1, 2, 3, 4) *
6	pcDNA3	0.166±0.008	(1, 2, 3, 4)*	0.196±0.02	(1, 2, 3, 4)*

* Significantly different from the groups considered in the parentheses according to the ANOVA test ($P<0.05$)

Table 2: Comparison of mean OD values of IgG1 determined by ELISA in the mice in two blood samplings

Number of group	Immunization regimen	IgG1 on day 42		IgG1 on day 63	
		Mean±SD	Sig ($P<0.05$)	Mean±SD	Sig ($P<0.05$)
1	pcSAG1+ pcROP2	0.181±0.015	---	0.157±0.013	---
2	pcGRA5+ pcSAG1	0.163±0.006	----	0.158±0.005	----
3	pcGRA5+ pcROP2	0.204±0.01	---	0.156±0.02	---
4	pcGRA5+pcSAG1+ pcROP2	0.173±0.007	---	0.173±0.007	---
5	PBS	0.168±0.003	----	0.168±0.001	----
6	pcDNA3	0.198±0.01	----	0.187±0.01	----

The highest values in the first and second blood samplings were related to pcGRA5+pcROP2 and pcDNA3 groups, respectively. However, the differences were not statistically significant ($P>0.05$). Moreover, the cut-off value determined according the IgG1

measurement in serum sample of seronegative mice was $0.168+3\times 0.03 = 0.177$ (Table 2).

According to Table 3, the lowest mean OD values in the first and second blood samplings were obtained for the pcDNA3 and PBS groups, respectively.

Table 3: Comparison of mean OD values for IgG2a determined using ELISA test in serum samples of mice for two blood samplings

Number of group	Immunization regimen	IgG2a on day 42		IgG2a on day 63	
		Mean±SD	Sig (<i>P</i> <0.05)	Mean±SD	Sig. (<i>P</i> <0.05)
1	pcSAG1+ pcROP2	0.252±0.01	(5, 6)*	0.240±0.03	(4,5,6)*
2	pcGRA5+ pcSAG1	0.258±0.01	(5, 6)*	0.282±0.01	(4,5,6)*
3	pcGRA5+ pcROP2	0.244±0.01	(5, 6)*	0.244±0.02	(4,5,6)*
4	pcGRA5+pcSAG1+ pcROP2	0.266±0.02	(5, 6)*	0.317±0.05	(1, 2, 3, 5, 6)*
5	PBS	0.149±0.002	(1, 2, 3, 4)*	0.148±0.006	(1, 2, 3, 4)*
6	pcDNA3	0.144±0.002	(1, 2, 3, 4)*	0.149±0.009	(1, 2, 3, 4)*

* Significantly different from the groups considered in the parentheses according to ANOVA test (*P*<0.05)

This is while the highest values in the first and second samplings related to pcGRA5+pcSAG1+pcROP2 group. Furthermore, the cut-off value obtained according to IgG2a measurement in serum sample of sero-negative mice was 0.155.

The lowest and the highest mean IFN- γ levels 72 hours after lymphocyte culture was re-

lated to the PBS and pcGRA5+pcSAG1+pcROP2 groups, respectively (Table 4).

The lowest IL-4 levels in mice 72 h after lymphocyte culture were related to the PBS group, while the highest level was related to the pcGRA5+pcSAG1+pcROP2 group, respectively (Table 4).

Table 4: Comparison of mean±SD IFN- γ and IL-4 levels determined by ELISA in samples of mice studied 72 hours after lymphocyte culture

Number of group	Immunization regimen	IFN- γ		IL-4	
		Mean±SD	Sig (<i>P</i> <0.05)	Mean±SD	Sig (<i>P</i> <0.05)
1	pcSAG1+ pcROP2	902±155.8	(4,5,6)*	52.0±17	(5,6)*
2	pcGRA5+ pcSAG1	1062±349	(5,6)*	38.4±8	(4, 5,6)*
3	pcGRA5+ pcROP2	892±196	(4,5,6)*	68±28.9	(5,6)*
4	pcGRA5+pcSAG1+ pcROP2	1278±136	(1,3,5,6)*	120±48	(2,5,6)*
5	PBS	167±6.6	(1,2,3,4)*	11±0.5	(1, 2, 3, 4)*
6	pcDNA3	180±4.6	(1,2,3,4)*	13±0.3	(1, 2, 3, 4)*

* Significantly different from the groups considered in the parentheses according to the Mann-Whitney test results (*P*<0.05)

Considering stimulation in the lymphocyte culture by the *Toxoplasma* antigen, the results obtained in the MTT assay for the experiment and control groups according to the stimulation index (SI), after the antigen stimulation in

the experiment and control groups, the highest and lowest SI values were obtained as 3.29 and 1.09 for the pcGRA5+pcSAG1+pcROP2 and pcDNA3 groups, respectively (Table 5).

Regarding the results obtained for mortality of the mice, mortality in the two control

groups; PBS and pcDNA3, started from days 4 and 5 after the challenge and all mice died until day 6.

Mortality in the pcGRA5+pcROP2 group started from day 6 after the challenge and all mice died until day 12. In the pcSAG1+pcROP2 group, the mortality of mice began from day 7 and all mice died until day 11. Considering the pcGRA5+pcSAG1+pcROP2 group, the mortality began from day 9 and all mice died until day 12. The survival rates of the mice in the days after parasitic challenge are illustrated in Fig. 3.

Table 5: Stimulation index (SI) in the MTT assay, after the lymphocyte culture stimulated with *Toxoplasma gondii* antigen in the experiment and control groups

Groups	SI
pcSAG1+ pcROP2	1.35
pcSAG1+pcGRA5	1.33
pcROP2+pcGRA5	1.36
pcGRA5+pcSAG1+pcROP2	3.29
PBS	1.00
pcDNA3	1.09

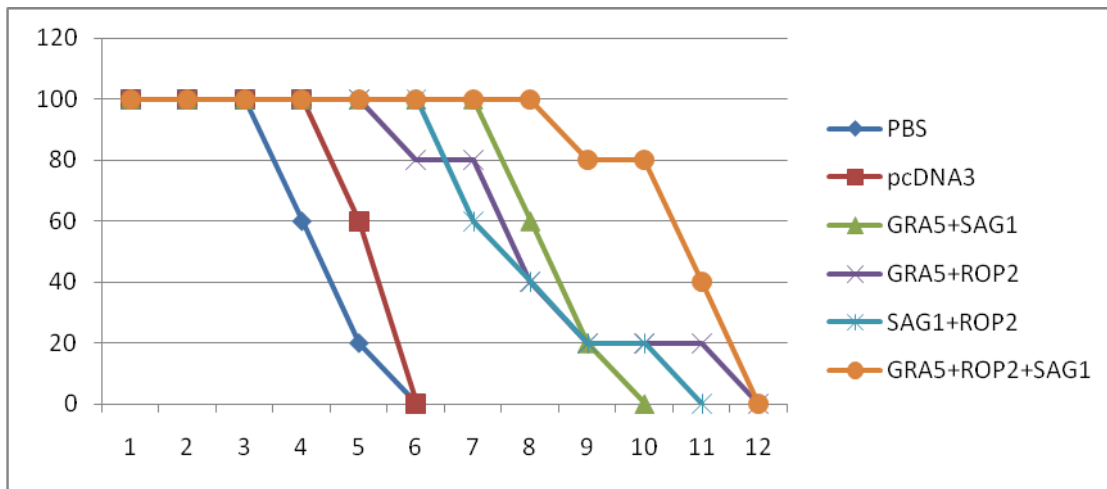


Fig. 3: Survival rates of immunized and control BALB/c mice after lethal challenge with 1×10^4 tachyzoite forms of *T. gondii* RH strain 7 weeks after the last immunization. Each group has five mice.

Discussion

Immunization with SAG1 and ROP2 proteins can lead to a widespread immune response, which is able to reduce the mortality rate of animals acutely contaminated by *T. gondii* and decrease the load of brain cyst in the infected animals (12) and the results of this study confirm the previous studies. However, the efficacy should be improved. Thus, we attempted to develop a novel vaccine by focusing on the genes encoding the major surface antigen (SAG1) (11, 17), as well as ROP2 (12, 15), and GRA5 (16).

Igarashi et al. have investigated the antigenic characteristics and production of recombinant GRA5, GRA7, and ROP2 proteins in *Toxoplasma* and reported the highest IFN- γ level in ROP2 plasmid. Accordingly, there was a possibility to employ these proteins in serological diagnosis or vaccination against the disease (14).

Dense granule antigens of *T. gondii* have been introduced as appropriate candidates for antiparasitic vaccines. Immunization with plasmids containing the afore-mentioned antigens, especially GRA5, leads to immune stimulation and antibody production (14, 18).

Gasior et al., for the first time, used complete recombinant *Toxoplasma* GRA5 antigen for serological diagnosis of human toxoplasmosis. Their study demonstrated that complete *Toxoplasma* rGRA5 was appropriate to be used as a component of cocktail antigen for identification of IgG antibody against toxoplasmosis (18).

Beyond its diagnostic value, GRA5 antigen has a protective role against the disease. Igarashi et al. investigated the mechanism of protection against brain cyst formation in BALB/c mice using the recombinant inhalant vaccine of rROP2, rGRA5, and rGRA7. They reported that immunization with the vaccine mentioned together with the cholera toxin resulted in relative protection in comparison with the control group and prevented development of *Toxoplasma* tissue cyst after oral drug administration (19).

Zenner et al. examined the subcutaneous vaccine of GRA2 and GRA5 antigens accompanied by FIA adjuvants for Fischer rats. The treatment was effective in preventing the disease spread from mother to baby. However, effectiveness of the approach for other animal models requires further investigations (20). In another study, GRA5, GRA7, and ROP2 genes from *T. gondii*, strain RH, were cloned and used for production of recombinant (14).

Considering the results obtained for survival of mice after being challenged by the RH strain showed that the lowest mean survival time in the PBS and pcDNA3 control groups were 126 and 138 h, respectively. This is while the mice that received recombinant plasmids had longer survival; such that the longest survival was observed in GRA5+SAG1+ROP2 and GRA5+SAG1 groups; 234 and 207 hours, respectively.

Administration of cocktail of *T. gondii* recombinant plasmids led to statistically significant enhancement of the survival rate of immunized mice.

In the first sampling, the total IgG titer in the groups that received recombinant plasmid did not have statistically significant difference

with that of the control groups. This is while in the second sampling, an increase was observed in the antibody level in the immunized groups, which was significantly different from the control groups ($P < 0.05$).

Conclusion

Simultaneous administration of two or three genes (cocktail vaccine) would enhance the expression of these genes. This, in turn would increase stimulation of the immune cells by the antigens and influences the humoral response and titer of the total IgG. Moreover, the results obtained from the ANOVA and Mann-Whitney tests indicated that with regard to the total IgG titer, the control groups were significantly different from all the groups immunized by recombinant plasmids.

Considering the results obtained on a daily basis for survival rate of the mice after challenge with the RH strain, it was observed that mortality in the control groups began from day 4, and all mice died until day 6 after the challenge. This is while in the GRA5+SAG1+ROP2 group, the mortality started from day 9 and all mice died until day 12.

According to the results obtained, although after challenge with live tachyzoite of *T. gondii*, strain RH, complete protection was not achieved in the immunized mice.

Acknowledgments

This study was supported financially by Iran National Science Foundation. The authors would like to thank the research department of Tarbiat Modares University, also Prof. Z.M. Hassan and Miss Ghasemi for their helpful aims. The authors declare that there is no conflict of interests.

References

1. Araujo FG. Immunization against *Toxoplasma gondii*. Parasitol Today. 1994; 10:358–360.

2. Couvreur G, Sadak A, Fortier B, Dubremetz JF. Surface antigens of *Toxoplasma gondii*. Parasitology. 1988; 97:1–10.
3. Capron A, Dessaint JP. Vaccination against parasitic diseases, some alternative concepts for the definition of protective antigens. Ann Inst Pasteur Immunol. 1988;139:109-117.
4. Cesbron MF, Capron A. Excreted-secreted antigens of *Toxoplasma gondii*: their origin and role in the host parasite interaction. Res Immunol. 1993;144:41-47.
5. Cesbron-Delauw MF. Dense granule organelles of *Toxoplasma gondii*: their role in the host parasite relationship. Parasitol Today. 1994; 10(8):293-296.
6. Saavedra R, Becerril MA, Dubeaux C, Lippens R, Vos MD, Herion P, Bollen A. Epitopes recognized by human T lymphocytes in the RoP2 protein antigen of *Toxoplasma gondii*. Infect Immun. 1996; 64:3858–3862.
7. Sadak A, Taghy Z, Fortier B, Dubremetz JF. Characterization of a family of rophtry proteins of *Toxoplasma gondii*. Mol Biochem Parasitol. 1998; 29: 203-211.
8. Velge-Roussel F, Marcelo P, Lepage AC, Buzoni-Gatel D, Bout D. Intranasal immunization with *Toxoplasma gondii* SAG1 induces protective cells into NALT and GALT compartments. Infec Immun. 2000; 68(2):969-972.
9. Nielsen HV, Lauemoller SL, Christiansen L, Buus S, Fomsgaard A, Petersen E. Complete protection against lethal *Toxoplasma gondii* infection in mice immunized with a plasmid encoding the SAG1 gene. Infect Immun. 1999; 67(12):6358-63.
10. Liu KY, Zhang DB, Wei QK, Li J, Li GP, Yu JZ. Biological role of surface *Toxoplasma gondii* antigen in development of vaccine. World J Gastroenterol. 2006; 2363:8-12.
11. Solhjo K, Ghaffari far F, Dalimi A, Sharifi Z. Enhancement of antibody immune response to a *Toxoplasma gondii* SAG1-encoded DNA vaccine by formulation with aluminium phosphate. J Med Sci. 2007; 7(3):361-367.
12. Hoseinian Khosroshahi K, Ghaffarifarf F, D'Souza S, Sharifi Z, Dalimi A. Evaluation of the immune response induced by DNA vaccine cocktail expressing complete SAG1 and ROP2 genes against *Toxoplasma gondii*. Vaccine. 2011; 29:778-783.
13. Holec-Gasior L, Kur J. *Toxoplasma gondii*: Recombinant GRA5 antigen for detection of immunoglobulin G antibodies using enzyme-linked immunosorbent assay. Exp Parasitol. 2010; 124(3): 272-8.
14. Igarashi M, Kano F, Tamekuni K, Kawasaki PM, Navarro IT, Vidotto O, et al. *Toxoplasma gondii*: cloning, sequencing, expression, and antigenic characterization of ROP2, GRA5 and GRA7. Genet Mol Res. 2008; 7(2): 305-13.
15. Hosseinian Khosroshahi K, Ghaffarifarf F, Sharifi Z, Dalimi A. Expression of complete rophtry protein 2 (ROP2) gene of *Toxoplasma gondii* in eukaryotic cell. Afr J Biotechnol. 2008; 7(24):4432–6.
16. Naserifar R , Ghaffarifarf F, Dalimi A, Sharifi Z. Cloning of *Toxoplasma gondii* Granular Antigen 5(GRA5) in Expression Eukaryotic Plasmid pcDNA3 and its Expression on CHO Cell.2011. J Ilam Univ Med Sci. 19(3):1-12.
17. Solhjo K, Ghaffari far, F, Dalimi A, Sharifi Z. Cloning and sequencing of *Toxoplasma gondii* major surface antigen (SAG1) gene. Arch Razi Inst. 2006; 61(2):73-79.
18. Holec-Gasior L, Kur J. *Toxoplasma gondii*: Recombinant GRA5 antigen for detection of immunoglobulin G antibodies using enzyme-linked immunosorbent assay. Exp Parasitol. 2010; 124(3): 272-8.
19. Igarashi M, Kano F, Tamekuni K, Machado RZ, Navarro IT, Vidotto O, et al. *Toxoplasma gondii*: Evaluation of an intranasal vaccine using recombinant proteins against brain cyst formation in BALB/c mice. Exp Parasitol. 2008;118(3):386-92.
20. Zenner L, Estaquier J, Darcy F, Maes P, Capron A, Cesbron-Delauw MF. Protective immunity in the rat model of congenital toxoplasmosis and the potential of excreted-secreted antigens as vaccine components. Parasite Immunol. 1999; 21(5):261-72.