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Original Article

Prevalence and Molecular Diagnosis of *Babesia ovis* and *Theileria ovis* in Lohi Sheep at Livestock Experiment Station (LES), Bahadurnagar, Okara, Pakistan

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Abstract

Background: *Babesia ovis* and *Theileria ovis* are among the important and main etiological agents causing ovine babesiosis and ovine theileriosis, causing severe economic losses among sheep and goats. The aim of the present study was to determine the prevalence and molecular diagnosis of *B. ovis* and *T. ovis* in Lohi sheep at Livestock Experiment Station Bahadurnagar, Okara, Pakistan.

Methods: The prevalence of *B. ovis* and *T. ovis* was investigated in 200 Lohi sheep of mixed age and sex by PCR during 2011. The assay was employed using primers Bbo-F & Bbo-R, specific for a 549-bp fragment in *B. ovis* genomic DNA and primers TSsr 170F & TSsr 670R, specific for a 520-bp fragment in *T. ovis* genomic DNA. The animals were also screened for both haemoparasites through stained thin blood smears.

Results: Thirty two (16%), 48 (24%) and 26 (13%) were the number of animals found positive for *B. ovis*, *T. ovis* and for mixed infection with both parasites respectively, through microscopy. Sixty eight (34%), 73 (37%) and 42 (21%) were the number of animals found positive for *B. ovis*, *T. ovis* and for mixed infection with both parasites respectively, through PCR test.

Conclusion: The results indicate the high sensitivity of PCR for surveying babesiosis and theileriosis and there is noteworthy prevalence of these diseases in sheep at an experimental station where environmental conditions are relatively controlled as compared to field conditions.

Introduction

In Pakistan, livestock contributed approximately 55.1% of the agriculture value added and 11.6% to the national GDP during 2011-2012 (1). Livestock sector's prospective role towards rural economic development can be gauged from the fact that 35-40 million rural populations are dependent on livestock, as every family has 2-3 cattle/buffaloes and 5-6 sheep/goats, thus leading to derive 30-40% of their income from these animals (2). Sheep and goats rearing carry tremendous importance in rural economy particularly for non-agricultural poor class of people (3). The population of sheep has been estimated as 28.4 million (M) in Pakistan and it yielded 0.037 M tons of milk for human consumption, 0.629 M tons of mutton and 0.043 M tons of wool during 2011-2012 (1). Punjab Province is the homeland for Lohi sheep and shares 24% of the total sheep population in the country (4).

Despite such a heavy population of livestock in Pakistan, the production by these animals is not as much as it could be. There are number of reasons for this discrepancy, e.g. poor breeding selection, management deficiencies and prevalence of blood born parasitic diseases (3). Pakistan, being a sub-tropical region of South Asia, has favorable environmental conditions for the growth of various ticks responsible for transmitting different haemoparasitic diseases in cattle and small ruminants. Small ruminant babesiosis and theileriosis develop in erythrocytes in peripheral blood and within the reticuloendothelial system.

Mostly three species of *Babesia* involved in causing the disease in ovine are; *B. motassi*, *B. crassa* and *B. ovis*, (5). The disease caused by *B. motassi* may be acute and/or chronic (6), whereas *B. crassa* appears to have little or no pathogenicity (7).

B. ovis is highly pathogenic especially in sheep and causes severe infection which is characterized by fever, anemia, icterus and

hemoglobinuria with mortality rates ranging from 30 to 50% in susceptible host during field infections (8). It leads to significant losses among small ruminants due to its drastic effect on hemobiotic system (9). *Babesia* species are also capable of causing diseases in humans (10) and is a significant public health threat due to emerging blood borne zoonosis in developing countries like Pakistan. *T. lestoquardi*, *T. seprata* and *T. ovis* are mostly involved among different species of *Theileria* that cause theileriosis in sheep. *T. lestoquardi* causes malignant theileriosis in sheep and goats where as *T. seprata* causes low or nonpathogenic theileriosis (11). *Theileria ovis* is an important and main etiological agent of ovine theileriosis, causing severe economic losses among sheep and goats in tropical and subtropical areas including Pakistan (12-15). The disease is manifested by fever, progressive weight loss, decrease in production and ultimately may lead to death.

Some workers studied the prevalence of both protozoans in areas which were different from the area which was selected for present study. Durrani et al. (12) has found *Theileria* spp. in 35% sheep blood samples in district Lahore. Iqbal et al. (13) has detected *B. ovis* in 50% sheep blood samples by PCR from southern Punjab (Pakistan). Similarly Durrani et al. (16) has detected *T. ovis* in 6% blood samples of sheep and goats through PCR, in Punjab and Khyber Pukhtoon Khaw Provinces of Pakistan. However, before present study, little or no work was done in Pakistan on molecular diagnosis of *Babesia* and *Theileria* species especially in Lohi sheep. These types of sheep maintained at Livestock Experiment Station (LES), Bahadurnagar, Okara showed clinical signs of Babesiosis and Theileriosis such as persistent fever, loss of body weight, jaundice, anemia and sudden death in young animals during the summer season in 2011- (May to July).

Traditionally the haemoparasitic diseases were diagnosed on the basis of clinical signs and microscopy. Both methods have the limitation of their inability to detect the subclinical (e.g. trophozoite stage of parasites in blood, which is very difficult to detect microscopically) and carrier animals. The carrier animals need to be brought in to limelight to know the exact status of haemoparasitic infestation in a herd so that disease control program can be implemented in a more rational way in order to enhance the productivity of the animals. The modern molecular tools of diagnosis have the ability to overwhelm the limitations of the conventional diagnostic techniques.

Keeping in view the advantage of PCR for identifying the exact species of *Babesia* and *Theileria* species and to know the prevalence of these haemoparasites, this study was initiated in Lohi sheep maintained at LES, Bahadurnagar, Okara. In Pakistan, the previous studies on haemoparasitic infestation in sheep were carried out in some other breeds of sheep. There was no such type of study conducted in Lohi sheep previously.

Materials and Methods

A total of 1360 Lohi sheep of different age groups and sex were maintained at LES, Bahadurnagar, Okara. During the summer season, from May to July 2011, clinical signs such as persistent high fever up to 105 °F, loss of body weight, jaundice, loss of appetite, anemia and sudden death were observed especially in young lambs (1-9 months old). During these four months, a total number of 200 blood samples were collected from the ear veins of 200 animals of different age groups and sex showing signs and symptoms of both haemoparasitic diseases. Thin smears were prepared, air dried, stained with Field's stain (HemaColor™, Merck), and examined under oil immersion on a microscope (Digi 3, Labomed, USA) for the presence of intracellular *Babesia*

schizonts and/or *Theileria* piroplasms based on morphology (17). At least 50 microscopic areas per slide were searched carefully. Those blood smears were recorded as negative for *Babesia* spp. or *Theileria* spp. in which no schizonts or piroplasms were observed.

DNA isolation and polymerase chain reactions

A 5-ml blood sample was also collected from jugular vein of each of the above described 200 sheep and placed in 10 ml clean sterile vacutainers containing ethylenediaminetetraacetic acid. They were brought to the laboratory and stored at -20 °C until used for DNA extraction for PCR amplification. Whole blood was used for the extraction of total genomic DNA using a commercially available DNA isolation kit (PureGene®, Gentra, Minnesota, USA) according to the manufacturer's instructions. The nucleic acid was extracted, also from known negative samples (40 healthy sheep maintained at a private Livestock farm at Okara). DNA concentration was determined spectrophotometrically at 260 nm and 280 nm and the number of genomic copies was determined by using Minevra Biolabs kit "VenorGem®" and the samples were stored at -20 °C till further use.

Three sets of primers (Table 1) were used for PCR amplification of different gene segments of both of the parasites. The first pair of oligo-nucleotide primer was used to amplify 549 base pair (bp) region of small sub unit rRNA (ssu rRNA) gene of *B. ovis* (18). The second primer pair specific for *Theileria* spp. was used for the amplification of small subunit ribosomal RNA (srRNA) gene fragment (1098 bp) from *Theileria* species genomic DNA as described by Allsopp et al. (19). The third primer pair, described by Altay et al. (20), was used to amplify SSU rRNA gene fragment of 520 bp length, which is specific for *Theileria ovis* genomic DNA. All PCR amplification reactions, including control negative samples, were carried out in a final volume of 25 µl containing 4 µl lysate or 2 ng purified DNA as

DNA template, 12.5 µl commercially available PCR master mix (Pyro Start™ Fast PCR Master Mix- 2X, Fermentas, Ontario, Canada), 2 U *Taq* DNA polymerase (Thermoscientific, California, USA) and 2 mM MgCl₂ (Thermo scientific, California, USA). The primers were used at a concentration of 10 pmol µl⁻¹ for *B. ovis* and 80 pmol µl⁻¹ for *Theileria* spp. and *T. ovis*.

PCR was carried out in a DNA thermal cycler (PEQLAB Biotechnologie, GmbH, Erlangen, Deutschland) with following cycling program for *B. ovis*: 94 °C for 3 min, followed by 35 cycles at 94 °C for 1 min, 62 °C for 1 min and 72 °C for 1 min, with a final extension step at 72 °C for 5 min. The amplification was carried with 30 cycles for *Theileria* species specific primer set. After initial denaturation at 94 °C for 3 minutes., each cycle involved denaturation for 1 min at 94 °C, annealing for 1

min at 60 °C and extension for 1 min at 72 °C, with a final extension step at 72 °C for 5 min. Cycling conditions for *T. ovis* were 96 °C for 3 min followed by 30 cycles at 94 °C for 30 s, 60 °C for 30 s and 72 °C for 2 min with a final extension step at 72 °C for 10 min. Ten microliters of the PCR products admixed with 6X DNA loading dye™ (Fermentas, Ontario, Canada) were sized by electrophoresis on a 1% agarose gel (Gene choice®, Maryland, USA) (1h at 90 V) with 1-kb ladder (Fermentas, Ontario, Canada) as size marker. The gels were stained with ethidium bromide (HP 47.1, ROTH, Karlsruhe) (250 µg/ml @ one drop/100ml of 1% agarose gel) and analyzed in a UV trans-illuminator (Dolphin -Doc, Wealtec, NV, USA) for visualization of PCR products.

Table 1: The primer pair name, sequence, product, target gene and annealing temperature of *Babesia ovis*, *Theileria* spp. & *Theileria ovis* primers used in the present study

PN	F.P. S. (5'-3')	PN	R. P.S (5'-3')	P (bp)	T.G	AT °C	Specificity	Reference
Bbo-F	TGGGCAGGAC- CTTGGTTCTTCT	Bbo-R	CCGCG- TAGCGCCGGCT AAATA	549	ssu rRNA gene	62	<i>B. ovis</i>	(18)
989 F	AGTTTCTGAC- CTATCAG	990 R	TTGCCITAAACT TCCTTG	1098	sr RNA gene	60	<i>Theileria</i> spp.	(19)
TSsr 170F	TCGAGAC- CTTCGGGT	TSsr 670R	TCCGGACATT- GTA AACAAA	520	SSU rRNA gene	60	<i>T. ovis</i>	(20)

PN=Primer Name, FPS=Forward Primer Sequence, RPS=Reverse Primer Sequence, P=Product, TG=Target gene, AT=Annealing Temperature

Results

The stained blood films revealed schizonts as teardrop-shape in pairs in red blood cells which were identified as *Babesia*, while piroplasm appeared in oval and ring forms, detectable inside the red blood cells and these were identified as *Theileria* (5, 17). Out of the 200 blood smears examined, 32 (16%) samples were positive for *Babesia*, 48 (24%) for *Theileria* while 26 (13%) were positive for mixed infections with both parasites through microscopy.

Data recorded for extracted DNA concentration by spectrophotometric analysis showed optical density values of 1.83 and 0.83 at 260 nm and 280 nm wavelengths, respectively. Sixty eight (34%) out of 200 samples showed amplification of a 549-bp fragment (Fig 1a) with the primer pair Bbo-F and Bbo-R, specific for *B. ovis*. Similarly 1098-bp fragment (Fig. 1b) was generated with primer pair 989F and 990R specific for *Theileria* spp. in 78/200 (39%) samples while a 520-bp fragment (Fig. 1c) was amplified with primer pair TSsr170F

and TSsr670R specific for *T. ovis* in 73/200 (37%) samples. Forty two (21%) samples showed mixed infection and were positive for both *T. ovis* and *B. ovis*. No such amplicon was detected in control negative samples. All samples positive with microscopic examination of blood smears, were also positive with PCR

test. The performance of both tests in terms of detection of positive cases has been compared in Table 2. Statistically (Z- test statistic), there was significant difference between the performance of PCR technique and microscopy for the diagnosis of haemoparasites.

Table 2: Comparison of prevalence of *T. ovis* & *B. ovis* in Lohi sheep by microscopic & PCR methods at LES, Bahadurnagar, Okara, Pakistan

Technique	Total No. of samples	No. of sheep positive for <i>B. ovis</i>	No. of sheep positive for <i>T. ovis</i>	No. of sheep positive for <i>T. ovis</i> & <i>B. ovis</i>
Microscopic Examination	200	32	48	26
PCR	200	68	73	42

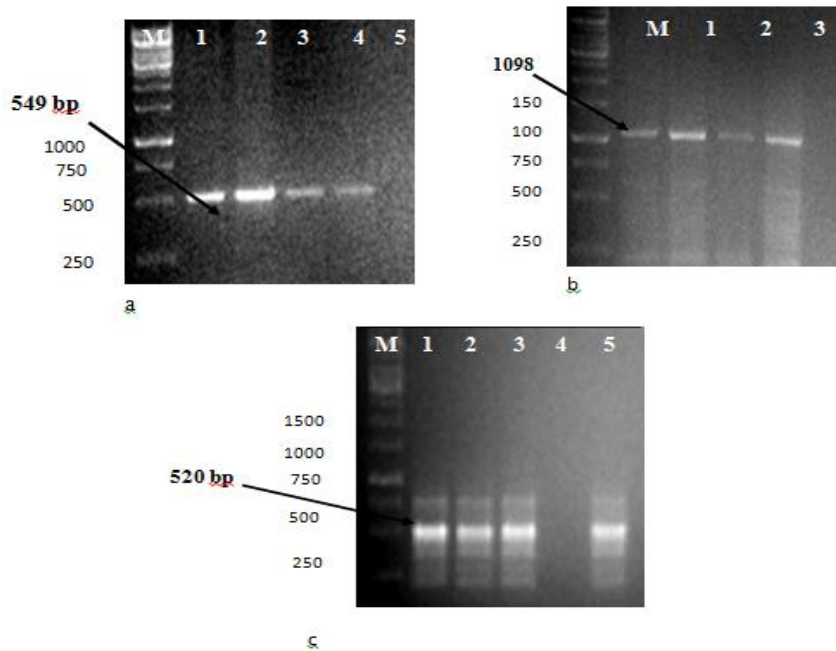


Fig. 1: Confirmation of *Babesia* and *Theileria* species by PCR amplification from Lohi sheep (a) Confirmation of *B. ovis* from field samples by PCR by amplification of four field samples with primer pair BboF and BboR at 549- bp. Lane M: Molecular Ladder; Lanes 1, 2, 3, 4: test samples + ve for *B. ovis*; Lane 5: Negative control (b) Confirmation of *Theileria* spp. from field samples by PCR by amplification of four field samples with primer pair 989 F and 990 R at 1098- bp. Lane M: Molecular Ladder; Lanes 1, 2, 3, 4: test samples + ve for *Theileria* spp.; Lane 5: Negative control (c) Confirmation of *T. ovis* from field samples by PCR by amplification of four field samples with primer pair TSsr 170F and TSsr 670R at 520- bp. Lane M: Molecular Ladder; Lanes 1, 2, 3, 5: test samples + ve for *T. ovis*; Lane 4: Negative control

Discussion

Several studies in Pakistan revealed that *B. ovis* and *T. ovis* are endemic in sheep and goats in different parts of the country (12, 13, 15, 16, 21, 22). There are very few case reports about sheep being infested with these parasites. This is the first molecular diagnostic report about the prevalence of *B. ovis* and *T. ovis* in Lohi sheep at LES, Bahadurnagar, Okara, Pakistan. Iqbal et al. (13) observed 50% prevalence of *B. ovis* in sheep in Southern Punjab through PCR which was higher as compared to the prevalence of *B. ovis* (34%) found during the present study. The difference in prevalence rate between the two studies may be attributed to the difference in geographical areas. Also the author of previous study has not mentioned the specific breed/s of sheep, which might be more susceptible to this disease as compared to Lohi breed. Durrani et al. (12) reported 27.5% (55/200) prevalence of *T. ovis* in sheep through PCR at district Lahore, which was less than the prevalence of the same protozoan (37%) in the present study. Although climatic conditions of the two study areas are almost the same, yet the higher prevalence of *T. ovis* in the present study cannot be compared with low prevalence reported in the previous study due to the facts that in present study the samples were collected in peak summer season which was ideal for tick infestation, as compared to the previous study in which the samples were collected in spring season, when the tick population was not as much as that was in summer season. Another reason for this discrepancy was the animal rearing type difference. In the present research, the samples were collected from intensive farming system at LES Bahadurnagar, Okara as compared to the collection of samples from individual animal keepings in the previous study. The prevalence for a mixed infection (21%) with *T. ovis* and *B. ovis* has also been observed during this study which is supported by a previous study conducted by Inci

et al. (23) who observed 1.7% (7/421) prevalence of mixed infection for *T. ovis* & *B. ovis* in sheep in central Anatolia (Turkey).

The Lohi sheep which is an indigenous breed in Pakistan is recognized as relatively disease resistant breed as compared to contemporary sheep breeds in this country. During this study, it was noted that this type of sheep is susceptible to tick born diseases. The environment at LES is relatively controlled as compared to the field conditions and tickicidals are being used regularly especially in the summer season. The results of this study are indicative of the fact that even a very few ticks can cause the spread of Babesiosis and Theileriosis. Although the disease prevalence recording in different age groups was not the main focus of this study, but it was observed that the disease was more prevalent in animals less than one year of age as compared to the animals having more than one year of age. This finding is in accordance to the findings by Ramzi et al. (24) and Iqbal et al. (13) who were of the view that probably the animals develop immunity against these parasites with the age. The disease was diagnosed in small ruminants through symptoms and microscopy in earlier studies in Pakistan (15, 21, 22). Microscopic detection methods are still the cheapest and fastest methods used to identify *Babesia* and *Theileria* parasites, although these have limited sensitivity and specificity. These techniques can detect for up to one infected erythrocyte per ten thousand cells, requiring the analysis of 100-200 fields, the equivalent to 0.5 μ l of blood. It is an inexpensive technique but requires an experienced person to differentiate species and is reliable only if the amount of parasites in the blood is high enough to be detected, which is usually only possible during acute cases (25). The observation of paired intra-erythrocytic merozoites is indicative of infection but there are other stages of the parasite like the trophozoites, which present different forms and sizes depending on the species, and these make their detection difficult and time consuming. An

exact differentiation between the parasites is crucial to understand their epidemiology. Detection methods based on nucleic acid identification and their amplification are the most sensitive and reliable techniques available today; they are fast, very specific and although most of them rely on sophisticated equipment (25). Recently, molecular techniques have become the preferred methods for diagnosis of babesiosis and theileriosis, because these techniques are more sensitive and specific than other conventional methods (20, 26).

In this study a molecular technique was used to diagnose parasitic infestation in Lohi sheep. Microscopy of blood smears once again proved to be less sensitive as compared PCR. Through the latter technique the parasites were identified and confirmed up to genus level. Morphologically only a few blood smears were identified up to species level. During this study, all the samples found positive for haemo-parasites through microscopy, were also found positive for these protozoan through PCR, however the molecular technique detected some additional cases (of Theileriosis) that were found negative with microscopy. The specificity of PCR product of these samples was confirmed when compared to the amplicon size of those samples which were found positive with both of the techniques. Five samples positive for *Theileria* infestation detected through PCR were not typed with *T. ovis* specific primers. The probable reason behind this might be the genome sequence difference (due to involvement of some other strain/s or species of *Theileria*) that needed additional primer pairs specific for them. This hypothesis can be confirmed by gene sequencing of SSU rRNA gene which was beyond the scope of this study. The results of present study are in agreement with the findings of previous studies (12, 13, 16) who also compared microscopic examination test with PCR in small ruminants and found PCR to be a sensitive method for *B. ovis* and *T. ovis* diagnosis. Similar observations have also been made earlier (18, 20, 27-32), while

comparing PCR with blood smear examination.

Conclusion

PCR for the detection of *B. ovis* and *T. ovis* is specific and sensitive. The tests is suitable for tracing carrier animals and provide a qualitative and validated measure that is useful in epidemiological surveys and follow ups for drug treatment in Lohi sheep. In addition, it would be useful while designing haemoparasitic (ovine babesiosis and ovine theileriosis) control program in endemic areas. The study also revealed that Lohi sheep is very much susceptible to babesiosis and theileriosis. The prevalence rate of haemoparasitic infestation in sheep at LES cannot be disregarded. Special attention should be paid for the control of tick infestation to the animals in order to enhance the livestock productivity.

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