

Tehran University of Medical Sciences Publication http://tums.ac.ir

Iran J Parasitol

Open access Journal at http://ijpa.tums.ac.ir

Iranian Society of Parasitology http://isp.tums.ac.ir

Original Article

Genetic Characteristics of *Echinococcus granulosus* from Fixed Paraffin-Embedded Tissue Samples in Human Isolates Based on the High-Resolution Melting Point Analysis in Sabzevar, Northeast Iran

*Hossein Elyasi¹, Tahmine Farbodnia², Ehsan Javaheri³

1. Cellular and Molecular Research Center, Sabzevar University of Medical Sciences, Sabzevar, Iran

2. Student Research Committee, Sabzevar University of Medical Sciences, Sabzevar, Iran

Received 20 Feb 2024 Accepted 27 May 2024

Keywords: Echinococcus granulosus; Genotyping; Hydatid cyst; High-resolution melting point analysis

*Correspondence Email: elyasihossein8@gmail.com helyasi2005@yahoo.com

Abstract

Background: There are ten genotypes of *Echinococcus granulosus* with different intermediate and final hosts affecting the parasite's life cycle and its transmission to humans. Therefore, this study was conducted to determine the genotype of isolated hydatid cysts using the simple and fast high-resolution melting point analysis (HRM) method.

Methods: The paraffin tissue samples of patients who underwent surgery were obtained from the pathology sample bank of Vasei and Emdad Hospitals in Sabzevar, Iran during 2010-2020. The DNA content of the samples was extracted after collecting and determining the characteristics using the DNA extraction kit. PCR was performed on the samples and the presence of the hydatid cyst genome was confirmed using the special Master Kit. Mix PCR of Solis Biodyne Company and Real-Time device (Bio-Rad) were used, and the genetic identity of hydatid cysts were determined.

Results: Out of 33 paraffin samples, 21 samples contained hydatid cyst DNA, two of which were from the brain and 19 from the liver tissues; 12 samples did not contain hydatid cyst DNAs. All liver samples were from sheep species (G1), and the brain samples were from buffalo species (G3). Therefore, 9.53% of the *Echinococcus* species collected were buffalo (G3), and 90.47% were sheep (G1) strain.

Conclusion: Based on previous patterns, HRM methods can be used for easy and quick identification of *Echinococcus* strains. The G1 strain was the dominant strain causing hydatid cyst in different human organs, including the liver and brain.



Copyright ${\rm ©}$ 2024 Elyasi et al. Published by Tehran University of Medical Sciences.

This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International license.

^{3.} Leishmaniasis Research Center, Sabzevar University of Medical Sciences, Sabzevar, Iran

⁽https://creativecommons.org/licenses/by-nc/4.0/). Non-commercial uses of the work are permitted, provided the original work is properly cited

Introduction

ystic echinococcosis (CE) or hydatid cyst is a chronic helminthic zoonotic disease distributed globally (1,2). This disease is widespread in sheep breeding countries or rural areas where animals are slaughtered unhygienically (3,4) in which CE or hematic cyst disease are endemic. Earlier studies, conducted in Iran, showed that hydatid cysts are common in sheep (5 to 72%), camels (11.4 to 70%), cows (3.5 to 38%), and goats (1.7 to 20%). Such human infections were continuously reported from different regions of the country (5). The organism causing this disease is the dog tapeworm or E. granulosus, which exists cyclically among carnivores and several herbivorous mammals (including sheep, camels, horses, cows, pigs, other mono-poisons, and wild herbivores). Humans are also considered as accidental intermediate hosts for the parasite. CE has significant economic and medical effects on livestock and humans in many parts of the world, including Iran (6).

This strain has a wide genetic diversity, and ten genotypes of different intermediate hosts have been reported for *E. granulosus* (G1-G10) so far, each of which is more compatible with a specific host. G1 is compatible with sheep, G2 Tasmanian sheep, G3 buffalo, G4 horse, G5 cow, G6 camel, G7 pig, G8 to G10 wild cycle (deer), and G9 milk strain; in addition, some of these strains have been reported in human cases (7).

Considering the lack of studying the genotype of human hydatid cyst in Iran and the importance of this disease to its population, the genotype of the disease-causing parasite was examined in the present study. In order to determine the parasite strain and to identify the main and intermediate hosts for better management and controlling this disease, the present study was designed to determine the molecular characteristics of human hydatid cyst strains in Sabzevar, Iran.

Materials and Methods

Area of Sampling

This study was conducted in Sabzevar, a northeastern city in Razavi Khorasan Province, Iran (Fig. 1). Its population is around 244,000 people; the area is a desert-like region with hot and dry climate at a longitude of 57° 42' 59.99" E and a latitude of 36° 11' 60.00" N. Its height above sea level is 978 meters, and the average annual rainfall is 191 mm, with an average annual temperature of 17.6 °C (8).

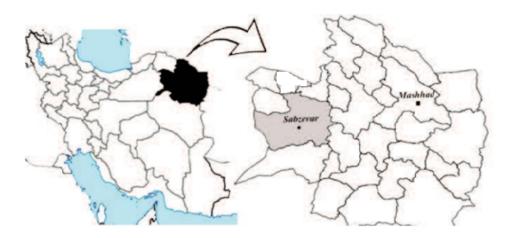


Fig. 1: The location of Sabzevar city in the map of Iran

Sampling

This study was carried out retrospectively during 2010 to 2020 in two major hospitals and medical training centers affiliated to Sabzevar University of Medical Sciences, Iran. The sample randomly included individuals who underwent surgery with a definitive diagnosis of the hydatid cyst disease.

Paraffin tissue samples of the patients, which were obtained from the bank of pathology samples at the laboratory of both Vasei and Emdad Hospitals of Sabzevar, Iran, were used. After collecting and determining the characteristics of the samples, they were transferred to the molecular cell laboratory at the medical school. After deparaffinization, the tissue inside the paraffin blocks was cut with a scalpel blade and placed inside a sterile microtube. Afterwards, 1000 microliters of xylenol were added to the tissue to dissolve the remaining paraffin completely. The numbered microtubes were then placed on the hot plate for 24 hours. The tissues were removed from the microtube and placed in a sterile one.

DNA extraction

The DNA extraction was done using Gent Bio's DNA extraction kit according to the manufacturer's protocol.

HRM assay

In order to identify *E granulosus* and determine the DNA sequence, a bio-Rad device with an HRM program was used. As control samples, the samples that were genotyped as well as the standard samples of certain genotypes such as G1, G3, and G6 were used to compare the analyses (9). The 300bp regions of the mitochondrial cox1 gene were commonly used to identify *Echinococcus* species (9). R/JB5 was amplified using primers (F/JB3 (5'-TTTTTGGGCATCCTGAGGTTTAT-3' and 5'-TAAAGAAAGAACATAATGAAAATG-3) (10).

In order to carry out the reaction in a volume of 20 microliters, first we prepared the samples together with the positive controls of *E. granulosus* species (G1, G3, G6) from Master Mix and primers and some sterile distilled water, and then we divided the volumes into microtubes. The predicted amount of DNA was added to the microtubes of samples and controls with the same concentration. After putting the lid on the microtubes, they were numbered and placed in the Real-Time device (Bio-Rad).

The thermal program was done for 10 seconds at 95 °C for denaturation, 15 seconds at 55 °C for annealing, and 10 seconds at 72 °C for final elongation. The cycle was repeated for 40 minutes for the replication of fragments.

In the current study, the genotype of hydatid cysts was investigated. The relationship between the genotype of hydatid cysts and the type of infected organ (e.g., liver, lung, kidney, and other organs) were also examined.

Ethical considerations:

The study was retrospectively conducted on tissue samples rather than human beings; the samples were obtained from the hospital bank of samples with the permission of the university research committee. Also, the study design was approved by the Ethics Committee at Sabzevar University of Medical Sciences, Iran (IR.MEDSAB. REC.1400.083).

Results

Overall, 33 samples were collected from two major hospitals in Sabzevar, Iran; two samples (from the liver tissue) were obtained from Vasei Hospital. Also, 31 samples were collected from Emdad Hospital, two belonging to the brain and 29 to the liver.

From these 33 paraffinated samples, DNA of hydatid cysts was extracted from 21 samples, while the DNA of hydatid cysts was not extracted from 12 samples. Two of the extracted samples were from brain and 19 from liver tissues. The G6 genotype were not identified in the studied samples, and 12 DNA samples were not extracted (Table 1).

No	Frequency	Sample type	Genotype	Percent	T_M	SD
1	19	Liver	G1	90.48	81.4	0.1
2	2	Brain	G3	9.52	81.9	0.07

 Table 1: Mean Melting point (TM), Standard Deviation (SD), and percentage of each genotype calculated for each genotype of *Echinococcus granulosus*

PCR melting curve results showed that the average melting temperature for G1 and G3 were 40.81 and 90.81 °C, respectively. The melting and HRM curves are shown in the normal and separation graphs in Fig. 2 and 3.

Fig. 2 shows the normalized melting analysis curves between the examined and standard samples. The normalized view of the graph is concerned with *E. granulosus* samples (G1 and

G3) in the temperature range of 80-84 °C. The HRM analysis of genes is determined by using standard samples determined in advance. On the other hand, melting analysis curves of G1 and G3 genotypes were used as standards to determine the samples. As can be seen, different genotypes are close to the known standard sample.

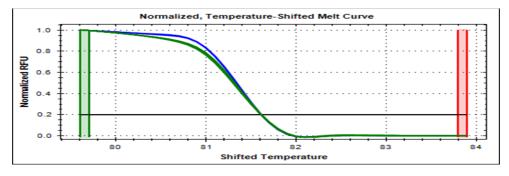


Fig. 2: The HRM analysis for samples in the normal graph

Fig. 3 shows the melting analysis curves separating the examined and standard samples. The HRM analysis of genes is determined by using pre-determined standard samples. Melting analysis curves of G1 and G3 genotypes have been used as standards to determine the samples. Hence, different genotypes are close to the known standard sample.

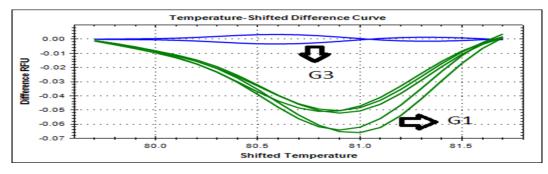


Fig. 3: The HRM analysis for samples in the discriminant graph

Color separation was used to identify the species in the charts. Therefore, the green col-

or was used in the G1 separation graph, and blue was used in the G3 separation graph.

Discussion

The present study was conducted to determine the genotype of isolated hydatid cysts using the HRM method in Sabzevar, a northeastern city in Iran. The study results primarily indicated that the genetic identity of hydatid cysts in a region could be determined with simple methods such as HRM methods. The dominant species in this region were G1 or the sheep species. In various studies conducted by researchers in different regions of Iran, other genotypes such as G1, G3 or G6 were also found (11-36) (Table 2).

No.	Frequency	Source	Area	Strains	Methods	Gene	References
1	50	Human (liver, lung,	Khorasan Razavi	G1-G3	RFLP	ITS1	11
		Spleen)		Senso			
		· · ·		strico			
2	22	Dog, Jackal (adult	Ilam	G1-G3	RFLP,	nad1	
		worm)			Sequencing		12
3	49	Domestic Animal	Central Province	G1, G2,	Sequencing	cox1	13
				G3			
4	47	Human (liver, lung, Spleen)	Tehran	G1, G3	Sequencing	cox1, nad1	14
5	60	Human (liver)	Mashhad	G1, G6	RFLP,	ITS1,	15
				,	Sequencing	cox1, nad1	
6	55	Human, Domestic	Ardabil	G1, G3	Sequencing	cox1, nad1	16
		animal					
7	43	Human	Busher	G1	Sequencing	cox1	17
8	9	Dog, Jackal (adult	Ardabil	G1	Sequencing	cox1, nad1	18
0	=-	worm)	TT 1	01.00	· ·	0.4	10
9	72	Human (liver)	Hamedan	G1, G3	Sequencing	Cox1,	19
10	•		Ŧ	<u> </u>	· ·	nad1	20
10	20	Human (liver)	Iran	G1, G3	Sequencing	nad5	20
11	122	Domestic Animal	North Khorasan	G1, G3,	Sequencing	cox1	21
				G6			
12	120	Domestic Animal,	Tabriz	G1	Sequencing	cox1	22
		Human					
13	15	Dog, Jackal (adult	Mazanderan	G1, G3	Sequencing	cox1	23
14	38	worm) Dog (adult worm)	Khorasan Razavi	G1	RFLP	ITS1	24
15	19	Human (liver, lung)	Yazd	G1, G6	Sequencing	cox1	25
16	8	Human (liver, lung)	Birjand	G1, G6	Sequencing	cox1, nad1	25
17	62	Domestic Animal,	Mazanderan	G1, G2,	Sequencing,	cox1, ITS1	20 27
17	02	Human	Mazanderan	G3, G1-G3	RFLP	cox1,1101	27
18	23	Human	Esfahan	G1-G3	Sequencing	cox1, nad1	28
19	4	Human	Tehran	G1-G3	Sequencing,	cox1,	29
		110111011	Tonnun	01 00	RFLP	nad1, ITS1	_>
20	55	Human	Azerbaijan	G1	RFLP	ITS1	30
21	29	Human	Tehran	G1, G3,	Sequencing	cox1, nad1	31
	_/		1 0111411	G6 G6		,	51
22	30	Human	Golestan	G1	RFLP	ITS1	32
23	11	Human	Ardabil	G1, G3	Sequencing	cox1, nad1	33
24	17	Human	Fars	G1, G6	Sequencing	nad1	34
25	1	Human	Kerman	G6	Sequencing	cox1, nad1	35
26	17	Domestic Animal	Sabzevar	G1	Sequencing	Cox1	36

Table 2: Published information concerning Echinococcus granulosus isolated in different regions of Iran

In a study by Sadri et al., the characteristics of hydatid cysts isolated during 2001-2002 from slaughtered animals in the industrial slaughterhouse of Yasouj, Iran was determined by PCR-RFLP method. Their research aimed to determine the molecular characteristics of hydatid cyst strains and their spread in the liver and lungs of animals slaughtered in the industrial slaughterhouse of Yasouj, Iran (37). In thir study, 93 animal hydatid cysts (31 sheep samples, 56 goat samples, and six cow samples) were collected from Yasouj industrial slaughterhouse; genomic DNA was extracted from the corresponding protoscolex by the standard phenol-chloroform method. They used PCR to amplify the rDNA-ITS1 fragment of each sample with EGR and EGF primers. The PCR products were first examined by electrophoresis and then cut with Alu I and Rsa I enzymes. The RFLP-PCR products of the said fragments were electrophoresed and investigated. The PCR method obtained the size of rDNA-ITS1 fragments of all liver and lung samples with similar bands and size of 1000bp. The pattern created using the RFLP test with AluI and RsaI enzymes showed the G1 genotype, the same sheep genotype of E. granulosus. The Yasouj study revealed that strain G1 was the dominant strain causing hydatid cyst disease in different organs of livestock, including the liver and lungs (37).

In the present study, a specific primer of the cox1 gene was used to identify different species of *E. granulosus*. Our results as presented in the graphs of the test samples' control samples (standard), showed the characteristics of the double-stranded DNA product based on its melting behavior. In addition, based on HRM PCR analysis, 19 samples of G1 genotype and two samples of G3 were identified from the total of 21 samples examined, where the G1 genotype was detected only in 19 sheep samples (90.48%), and G3 genotype was detected only in 2 buffalo samples (9.52%). However, the G6 genotype was not detected in our samples. Therefore, the pre-

dominant genotype found in Sabzevar, Iran was the G1 genotype. Therefore, our findings are consistent with the above-mentioned study conducted in Yasouj, Iran (37), even though the study methods were different.

In another study, 46 hydatid cyst samples, 18 obtained from female and 28 from male cases, were subjected to molecular analysis (38). Then, the samples were identified using PCR and sequencing methods, where all the samples showed G1 or sheep genotype in humans (38). Still another study conducted in southwest Iran examined 334 hydatid cyst samples collected from the liver and lungs of 141 sheep, 104 calves, 84 goats, and 5 human samples (39). DNA extraction and PCR of the ITS1 fragment were done. Then RFLP was performed and fragments of the ITS1 gene were sequenced. Their results showed that the G1 genotype was present in all human, bovine, goat and sheep samples but the camel strain (G6) was not present among the samples. In line with the present study, their molecular findings also indicated that the dominant genotype in southwestern Iran was the sheep strain (G1), which is common among human populations, cattle, sheep, and goats (39).

In Kania region of Turkey during 2018-2019, after DNA extraction and sequencing of 83 cysts (57 bovine and 26 human cysts), 82 cysts with Sensu strict genotype (G1-G3) were identified together with several human cysts as G4 (equinus) (40). Moreover, Shafiei et al., examined the relationship between parasite genotype and hydatid cyst location, in which they found that some liver cysts had a G1 genotype. In contrast, six cysts related to the peritoneum, kidney and brain were G2, but one of the pelvic cysts was identified as G3 (41).

Moreover, Hedayati and colleagues collected 47 paraffin blocks related to patients with hydatid cysts in Mazandaran, the coastal province to the Caspian Sea in the north of Iran. After deparaffinization, PCR, and sequencing, it was determined that 66.6% of the samples had G1 genotype while 33.3% had G1 genotype, which were related to G2-G3 genotypes (42).

Conclusion

All liver samples were G1, and all brain samples were G3. Therefore, 9.52% of the *Echinococcus* species collected were G3 (buffalo species), while 90.48% were G1 (sheep species). The predominant strain of the *E. granulosus* parasite in Sabzevar, a city located to northeast of Iran, was the same as other reports from other parts of Iran. It is worth mentioning that this parasite is the main host in sheep and dogs, and the intermediate hosts are the cattle.

Acknowledgements

The authors of this study would like to express their gratitude to the Deputy of Research and Technology at Sabzevar University of Medical Sciences, Sabzevar, Iran for their approval and financial support.

Conflict of interest

The authors declare that there is no conflict of interest.

References

- 1. Eckert J, Deplazes P. Biological, epidemiological, and clinical aspects of echinococcosis, a zoonosis of increasing concern. Clin Microbiol Rev. 2004;17(1):107-35.
- 2. Moro P, Schantz PM. Echinococcosis: a review. Int J Infect Dis. 2009;13(2):125-33.
- Murray CJ, Vos T, Lozano R, et al. Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet. 2012;380(9859):2197-223.
- 4. Thompson RC. Biology and Systematics of *Echinococcus*. Adv Parasitol. 2017;95:65-109.

- Haniloo A, Farhadi M, Fazaeli A, Nouri, et al. Genotype characterization of hydatid cysts isolated from Zanjan using PCR-RFLP technique. J Adv Med Biomed Res. 2013;21(84):57–65.
- 6. Abbasi I, Branzburg A, Campos-Ponce M, et al. Copro-diagnosis of *Echinococcus* granulosus infection in dogs by amplification of a newly identified repeated DNA sequence. Am J Trop Med Hyg. 2003;69(3):324-30.
- Nakao M, McManus DP, Schantz PM, et al. A molecular phylogeny of the genus *Echinococcus* inferred from complete mitochondrial genomes. Parasitology. 2007;134(Pt 5):713-22.
- Zamand S, Alidadi H, Najafpoor A, Taghavimanesh V, Akbari Sharak N, Najafi Saleh H. Data on trend changes of drinking groundwater resources quality in Sabzevar city (A case study). Data Brief. 2018; 20: 889-893.
- Rostami S, Shariat Torbaghan S, Dabiri S, et al. Genetic characterization of *Echinococcus* granulosus from a large number of formalinfixed, paraffin-embedded tissue samples of human isolates in Iran. Am J Trop Med Hyg. 2015;92(3):588-94.
- Metwally DM, Qassim LE, Al-Turaiki IM, et al. Gene-based molecular analysis of COX1 in *Echinococcus granulosus* cysts isolated from naturally infected livestock in Riyadh, Saudi Arabia. PLoS One. 2018;13(4):e0195016.
- Borji H, Bahar MM, Naghibi A, et al. Genotyping of human cystic echinococcosis in northeastern Iran. Ann Parasitol. 2018;64(4):323-329.
- 12. Dalimi A, Shamsi M, Khosravi A,et al. Genotyping *Echinococcus granulosus* from Canine Isolates in Ilam Province, West of Iran. Iran J Parasitol. 2017;12(4):614-621.
- Abedi B, Maghsood AH, Khansarinejad B, et al. Genotyping of *Echinococcus granulosus* isolates from livestock based on mitochondrial cox1 gene, in the Markazi province, Iran. J Parasit Dis. 2019;43(4):592-596.
- 14. Nikmanesh B, Mirhendi H, Ghalavand Z, et al. Genotyping of *Echinococcus granulosus* Isolates from Human Clinical Samples

Based on Sequencing of Mitochondrial Genes in Iran, Tehran. Iran J Parasitol. 2014;9(1):20-7.

- Berenji F, Shamsian SA, Nouri Daloee M, et al. Genotyping of *Echinococcus granulosus* Isolates from Human in Khorasan Province, North-Eastern Iran. Iran J Parasitol. 2019;14(1):52-58.
- 16. Pezeshki A, Akhlaghi L, Sharbatkhori M, et al. Genotyping of *Echinococcus granulosus* from domestic animals and humans from Ardabil Province, northwest Iran. J Helminthol. 2013;87(4):387-91.
- Babaei Z, Taherkhani R, Barazesh A, et al. Genotyping and phylogenetic analysis of hydatid cysts isolated from livestock in Bushehr province, Iran. J Parasit Dis. 2021;45(1):197-203.
- Zarei, Z., Ghalehbin, B.M., Akhoundi, B. et al. Genotyping of *Echinococcus granulosus* isolated from canine in Northwest Iran. J Parasit Dis. 2023; 47(4):757–761.
- Shahbazi AE, Saidijam M, Maghsood AH, et al. Genotyping of Fresh and Paraffinized Human Hydatid Cysts Using *nad1* and *cox1* Genes in Hamadan Province, West of Iran. Iran J Parasitol. 2020;15(2):259-265.
- Shahabi S, Sarkari B, Barazesh A. *Echinococcus granulosus* sensu stricto G1 is the predominant genotype in human and livestock isolates from Turkey and Iran, based on mitochondrial nad5 gene differentiation. Parasit Vectors. 2021;14(1):369.
- Salehi M, Yaghfoori S, Bahari P, et al. Molecular Characterization of *Echinococcus* granulosus Sensu Lato from Livestock in North Khorasan Province, Iran. Iran J Parasitol. 2018;13(4):577-586.
- 22. Zarrabi Ahrabi S, Madani R, Shemshadi B, et al. Genetic Affinity of *Echinococcus* granulosus protoscolex in Human and Sheep in East Azerbaijan, Iran. Arch Razi Inst. 2020;75(1):47-54.
- 23. Gholami S, Jahandar H, Abastabar M, et al. *Echinococcus granulosus* Sensu Stricto in Dogs and Jackals from Caspian Sea Region, Northern Iran. Iran J Parasitol. 2016;11(2):186-194.
- 24. Ghabdian S, Borji H, Naghibi A. Molecular identification of *Echinococcus granulosus* strain

in stray dogs from Northeastern Iran. Vet Parasitol Reg Stud Reports. 2017; 9:6-8.

- Kargar S, Dalimi A, Eslami G, et al. Cystic Echinococcosis in Central Iran: G1 and G6 Genotypes in Patients. Surg Infect (Larchmt). 2022; 23(5):451-457.
- Karamian M, Haghighi F, Hemmati M. Genotyping of Cystic Hydatidosis Agents in Birjand, Eastern Iran. Mod Care J. 2017;14(3):e64531.
- 27. Gorgani-Firouzjaee T, Kalantrai N, Ghaffari S, et al. Genotype characterization of livestock and human cystic echinococcosis in Mazandaran province, Iran. J Helminthol. 2019;93(2):255-259.
- 28. Sharbatkhori M, Mirhendi H, Jex AR, et al. Genetic categorization of *Echinococcus* granulosus from humans and herbivorous hosts in Iran using an integrated mutation scanning-phylogenetic approach. Electrophoresis. 2009;30(15):2648-55.
- 29. Zhang L, Eslami A, Hosseini SH, et al. Indication of the presence of two distinct strains of *Echinococcus granulosus* in Iran by mitochondrial DNA markers. Am J Trop Med Hyg. 1998;59(1):171-4.
- Vahedi A, Mahdavi M, Ghazanchaei A, et al. Genotypic characteristics of hydatid cysts isolated from humans in East Azerbaijan Province (2011-2013). J Anal Res Clin Med. 2014; 2(3):152-157.
- 31. Nikmanesh B, Mirhendi H, Ghalavand Z, et al. Genotyping of *Echinococcus granulosus* Isolates from Human Clinical Samples Based on Sequencing of Mitochondrial Genes in Iran, Tehran. Iran J Parasitol. 2014;9(1):20-7.
- 32. Gholami Sh, Sosari M, Fakhar M, et al. Molecular Characterization of *Echinococcus* granulosus from Hydatid Cysts Isolated from Human and Animals in Golestan Province, North of Iran. Iran J Parasitol. 2012;7(4):8-16.
- Pezeshki A, Akhlaghi L, Sharbatkhori M, et al. Genotyping of *Echinococcus granulosus* from domestic animals and humans from Ardabil Province, northwest Iran. J Helminthol. 2013;87(4):387-91.
- 34. Sadjjadi SM, Mikaeili F, Karamian M, et al. Evidence that the *Echinococcus granulosus* G6

genotype has an affinity for the brain in humans. Int J Parasitol. 2013;43(11):875-7.

- 35. Hajialilo E, Harandi MF, Sharbatkhori M, et al. Genetic characterization of *Echinococcus* granulosus in camels, cattle and sheep from the south-east of Iran indicates the presence of the G3 genotype. J Helminthol. 2012;86(3):263-70.
- Elyasi H, Golmohammadi R. Genetic Identification of *Echinococcus granulosus* Isolates from Domestic Animals in Sabzevar, Northeast Iran. Iran J Parasitol. 2020;15(4):621-623.
- 37. Sadri A, Moshfe A, Doosti A, et al. Characterization of Isolated Hydatid Cyst from Slaughtered Livestock in Yasouj Industrial Slaughterhouse by PCR-RFLP. Armaghan Danesh, 2012; 17(3): 243-252.
- Ergin S, Saribas S, Yuksel P, et al. Genotypic characterization of *Echinococcus* granulosus isolated from human in Turkey. Afr J Microbiol Res. 2010; 4(7):551–555

- Khademvatan S, Yousefi E, Rafiei A, et al. Molecular characterization of livestock and human isolates of *Echinococcus granulosus* from south-west Iran. J Helminthol. 2013;87(2):240-4.
- 40. Macin S, Orsten S, Samadzade R, et al. Human and animal cystic echinococcosis in Konya, Turkey: molecular identification and the first report of *E. equinus* from human host in Turkey. Parasitol Res. 2021;120(2):563-568.
- 41. Shafiei R, Ghatee MA, Jafarzadeh F, et al. Genotyping and phylogenetic analysis of unusually located hydatid cysts isolated from humans in north-east Iran. J Helminthol. 2019;94:e64.
- 42. Hedayati Z, Daryani A, Sarvi S, et al. Molecular Genotyping of the Human Cystic Echinococcosis in Mazandaran Province, North of Iran. Iran J Parasitol. 2019;14(1):151-158.